DYNAMIC MODELING OF THE ACTIVATED SLUDGE PROCESS FOR REAL TIME MONITORING AND CONTROL

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ABSTRACT

In this paper, a dynamic model of the activated sludge process is formulated. The model is implemented in a modular hierarchical structure. Different biological reactions for COD removal, nitrification and denitrification are arranged in cascade and are active depending on the operation conditions. The implementation suits real time monitoring and control of the process. The implementation is configured to a very large plant to study its possible control strategies.

KEYWORDS

Virtual Instruments, Gabal El Asfar Wastewater Treatment Plant.

INTRODUCTION

The ever more stringent environmental and health regulations together with the demand of cost effective plants have made the monitoring, control and automation of wastewater treatment process a top priority. Distributed Computer Systems (DCS) have been successful on many different types of industrial processes and it has started to appear in wastewater treatment plants. Irrespective of the progress of sensor technology, certain measurements remain difficult to be made by automatic means offering the continuity and the reliability that is so important in the automatic control of certain treatment processes (Vanrolleghem and Lee, 2003).

When the main process mechanisms are known, the mathematical presentation and the dynamic modelling of physical, chemical and biochemical processes can be applied off-line to simulate the plant performance. Dynamic modelling is very advantageous if applied to real time monitoring and control of treatment processes. However for real time use, more research work is required. In (Zaher, 2000), a new complex model was built and implemented in a very flexible way to be suitable for real time monitoring and control of the Gabal El Asfar Wastewater Treatment Plant (1.0 million m^3/day), the biggest treatment plant in the Middle East. This paper presents the developed model and its implementation.

PROCESS MODEL

The Activated Sludge Process is very common in secondary treatment of domestic wastewater. In the plant under consideration, the AS process consists of two main components, the Aeration Tank (A.T.) and the final settling tank (Final Clarifier F.C). Both components are considered in the model.

Aeration tank modeling

The main biological mechanisms in the activated sludege process are the utilization of substrate and the concurrent growth of organisms (Bailey and Ollis, 1986). Non-viable biomass degrades a considerable part of substrate through enzyme kinetics (Jones, 1973). For all reactions 2 kinetic relationships are considered. The first is an enzyme-accelerated reaction and it is associated with non-viable biomass (not growing). The second is Monod growth and it will be associated with viable biomass (growing). Thus, 6 bacterial populations will be described within the model. They comprise three types and each type splits into viable and nonviable. Heterotrophs are responsible for COD removal. Facultative heterotrophs are responsible for denitrification and present the portion of heterotrophs that are able to utilise

nitrate oxygen when the dissolved oxygen is very low. Autotrophs are responsible of oxidising ammonium to nitrate. For all the species Michaelis – Menten kinetic relationship will represent the enzyme kinetics and Monod kinetics will represent the growth. Both will use double term definitions. One term represents the substrate uptake and another term for the uptake of DO (nitrate in the case of denitrification). Michaelis - Menten enzyme reaction rate is:

$$\psi(t) = \psi_{\max}\left[\frac{S(t)}{K_m + S(t)}\right] \left[\frac{DO(t)}{K_o + DO(t)}\right]$$
(1)

Similarly, the Monod term for growth rate:

$$\mu(t) = \mu_T \left[\frac{S(t)}{K_s + S(t)} \right] \left[\frac{DO(t)}{K_o + DO(t)} \right]$$
(2)

The growth rate is adjusted for temperature using the following empirical equation:

$$\mu_T = \mu_{\max} \times 1.07^{(T-20)} \tag{3}$$

To maintain the hierarchical structure the mass balance is applied for each component (defined in the nomenclature).

$$\frac{dX_{v}}{dt} = \frac{F(t)}{V} (X_{v,in} - X_{v}) + \mu(t)X_{v} - K_{d}X_{v}$$
(4)

$$\frac{dX_{nv}}{dt} = \frac{F(t)}{V} (X_{nv,in} - X_{nv}) + K_d X_v$$
(5)

$$\frac{dS}{dt} = \frac{F(t)}{V} (S_{in} - S) - \frac{\mu(t)}{Y_x} X_v - \psi(t) X_{nv}$$
(6)

$$\frac{dDO}{dt} = C(\frac{F(t)}{V}(DO_{in} - DO) + F\frac{DOC}{V} - mm\frac{X_{i}}{1000}) - KDO(\frac{\mu(t)}{Y_{x}}X_{v} + \psi(t)X_{nv})$$
(7)

For the components X_i that are not taking part in the reaction the following balance equation should be applied:

$$\frac{dX_{i}}{dt} = \frac{F(t)}{V} (X_{i,in} - X_{i}) - \frac{\mu(t)}{Y_{x}} X_{v} - \psi(t) X_{nv}$$
(8)

For the flow arrangement a series of mixed tanks where:

$$\left(Unit_{input}\right)_{i=1 \text{ to } n} = \left(Unit_{output}\right)_{i=i-1 \text{ to } n} \text{ Number of surface areators} = n+1$$
(9)

For the units that receives the returned flow and sludge, inlet concentrations are given by: $C_i = (F(t) C_i + f_R C_R) / (F(t) + f_R)$ (10)

Oxygenation Intensity is adjusted due to temperature $F_T = 1.02^{(T-20)}$ and due to dissolved salts Z $F_{DS,DO} = \left(\frac{475 - 2.65Z/1000}{33.5 + T} - DO\right)/8.8 \rightarrow F = F_T * F_{DS,DO}$ (11)

Final clarifier modeling

According to the work of (Takacs *et. al.*, 1991) the final clarifier is divided to horizontal layers. Equation 12 apply the flux theory to estimate each layer concentration:

$$\frac{dX}{dt} = \frac{F(t)}{V} (X_{in} - X) + \frac{A}{V} (V_{s1} X_{above} - O \quad V_{s2} X)$$
(12)

Inputs to equation (12) should be alternated according the layer position from the feed point as follows:

-When the current layer is at the feed point, flow F is assigned the total flow value, i.e. influent plus return sludge flow, and Xin is the entering MLSS.

-When the current layer is above the feed point, flow F is assigned the influent flow value and if it is the top layer, $X_{above} = 0$.

-When the current layer is beneath the feed point, flow is assigned the under flow value, i.e. return sludge flow plus waste sludge flow

- For all layers O=1 except the bottom O=0.

Where v_{sI} , v_{s2} in equation (12) are the settling velocities of two subsequent layers calculated by : $v_s = \min(v_{\max}, v_0[e^{-b_1 \cdot X} - e^{-b_2 \cdot X}])$ (13)

Where the parameters v_{max} , v_0 , b_1 and b_2 can be determined from the following empirical equations (Catunda and VanHaandel, 1992; Takacs *et. al.*, 1991):





Figure 1 Model tree diagram



MODEL IMPLEMENTATION

The previous formulation of the process model enables the model implementation using data driven programming. Thus, using a reasonable time step each variable can be calculated without the need to implement numerical solution routines. The results are very similar to solving the equation using the Euler numerical method. Equations from (1) through (17) are implemented on LabView, instrumentation-programming package, as virtual instruments (VIs). Thus, they can be linked to real instruments and actuators of the plant to function on-line. The model parameters can be tuned on-line. The process can be monitored on real time. Different control strategies can be tested and applied to either directly sensed variables or those calculated by the model.

Figure (1) shows the hierarchical structure of the implementation. Three biological processes are considered: the COD removal, the nitrification and the denitrification. They are in a cascade structure. Each process will be activated according the DO level: (< 0.2) for denitrification in the anoxic zones, (> 0.2) for COD removal and (> 1) for nitrification to be activated. Figures 2 and 3

show user interfaces to the process units. Through one window, the plant operator can monitor the evolution of different components in the aeration tank by just pressing the appropriate buttons. The components are calculated by the model. The application can be easily extended to have some of the components measured on-line, validated and the correct value shown. The solids profile in the final clarifier is viewed and can be checked regularly by inspecting the final clarifier unit or compared to turbidity measurements. A friendly interface to the process controls was developed (not shown). The plant operator can act or adjust the set points manually and the results will be reflected directly on the other interfaces. Also, control actions can be automated in relation to the estimated/measured variables and just viewed through the interface.

CONCLUSIONS

The presented model applies the principal kinetics of RAS biological reactions, mass balances each component, and empirically formulates major disturbances (temperature, dissolved salts and settleability characteristics). It is implemented on an instrumentation-programming package in a hierarchical modular structure. Different biological processes are sorted in cascade where they will be activated according the practical DO level. The modular structure and the use of instrumentation programming make the implementation easy to be extended and connected to sensors and actuators. Indeed, the implementation uses a friendly interface The model parameters can be estimated off-line. Different control strategies and operation scenarios can be studied. After tailoring the instrumentation and control equipment suitable to the plant, the model can be regularly checked and recalibrated as it is in use and continue for real time monitoring and control of the AS plant.

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NOMENCLATURE

μ (t) : bacterial growth rate (d ⁻¹)	Ko: half-saturation coefficient for Oxygen.
Ψ (t): rate of enzyme reaction (d ⁻¹)	Ks: half-saturation coefficient for substrate
μ_{max} : maximum specific growth rate (d ⁻¹)	Mm: Oxygen required by endogenous respiration mgO_2/g
Ψ max: the maximum enzyme rate (d ⁻¹)	biomass.
C: mass balance parameter	S: substrate concentration (mg/l).
C: Influent concentration (mg/l)	Sin: influent substrate concentration (mg/l).
$C_{\mathbb{P}}$: Return concentration (mg/l)	SSVI: stirred sludge volume index
DO(t): current dissolved oxygen concentration	T: Temperature °C.
(mg/l).	V(t) : reactor volume (m ³).
DOC(t) :oxygenation intensity in mg O ₂ /min.	Xnv: non viable bacteria concentration (mg/l).
Doin(t): DO concentration in the influent flow	Xt : total bacterial cells concentration (mg/l).
(mg/l).	Xv: viable bacteria concentration (mg/l).
F(t): influent flow (m ³).	Xv,in: influent viable bacteria (mg/l).
$F_{\rm p}$: return flow (m ³)	Yx: the bacterial cells yield factor (mg/l).
Kd: death rate (d^{-1}) .	Z: Dissolved salts concentration (mg/l).
KDO: mg COD equivelent of mg mass of	
substrate	